

# Composition of the essential oils of *Nepeta ispahanica* Boiss. and *Nepeta binaludensis* Jamzad from Iran

Abdolhossein Rustaiyan<sup>1\*</sup> and Kobra Nadji<sup>2</sup>

<sup>1</sup>School of Pharmacy, Shahid Beheshty University of Medical Science, PO Box 14155-6153, Tehran, Iran

<sup>2</sup>School of Pharmacy, Tehran University of Medical Science, Tehran, Iran

Received 22 March 1997

Accepted 12 May 1998

**ABSTRACT:** The composition of the essential oils of *Nepeta ispahanica* Boiss. and *N. binaludensis* Jamzad was investigated by means of GC, GC–MS and <sup>1</sup>H-NMR spectra of the main compounds. 1,8-Cineole was the most abundant component in both oils: *Nepeta ispahanica* (66%) and *N. binaludensis* (42%). However, nepetalactone (25%), linalol (4%),  $\alpha$ -terpineol (4%) and  $\beta$ -pinene (3%) were major constituents of *N. binaludensis*. The structures of 1,8-cineole, nepetalactones and  $\beta$ -pinene were confirmed by their <sup>1</sup>H-NMR spectra. Copyright © 1999 John Wiley & Sons, Ltd.

**KEY WORDS:** *Nepeta ispahanica* Boiss.; *Nepeta binaludensis* Jamzad; Lamiaceae; essential oil; 1,8-cineole; nepetalactone

## Introduction

The genus *Nepeta* (Lamiaceae), with almost 280 species, is widespread in Europe, Asia and in a few parts of Africa.<sup>1</sup> Some species are used in folk medicine, *N. cataria* (catnip), for instance, is used as a fortifier, a disinfectant and cure against colds.<sup>2</sup> Bacteriostatic and fungistatic properties of the essential oil from *N. cataria* have been investigated by F. Perineaw *et al.*<sup>3</sup>

Chloroform extracts of *N. kopetdaghensis* were found to be active as a bacteriostatic, a diuretic and a cure against eczema-type skin disorders.<sup>4</sup> Alcoholic preparations of *N. hindostana* decreased the level of serum lipids and lipoproteins, and might thus supplement remedies against arteriosclerosis.<sup>5</sup>

Many *Nepeta* species contain the diastereomeric nepetalactones, substituted cyclopentanoid iridodial derivatives,<sup>6</sup> which are known as powerful attractants for cats.<sup>7</sup>

## Experimental

The aerial parts of *N. ispahanica* Boiss. growing wild in Roodshoor, between Saveh and Tehran in north-west Iran at an altitude of 1900 m, were collected in July 1993 (voucher No. AR, 129) and the aerial parts of *N. binaludensis* Jamzad. growing wild on Binalud mountain (2300 m elevation, province Khorassan in north-east

Iran) were collected in July 1993 (voucher No. AR, 130).

Vouchers are deposited at the Herbarium of the Department of Botany, Shahid Beheshty University, Tehran, Iran.

## Isolation of the Essential Oils

The aerial parts of the two species were ground and the essential oils isolated by hydrodistillation for 4 h, using a Clevenger-type apparatus. After decanting and drying over anhydrous sodium sulphate, the corresponding yellowish coloured oils were recovered.

## Gas Chromatography and GC–MS

GC analysis was performed using a Packard 439 chromatograph equipped with a CP Sil 5CB column, (25 m × 0.25 mm i.d., film thickness 0.39  $\mu$ m), oven temperature programme from 60°C at 5°C/min to 220°C. N<sub>2</sub> was used as carrier gas at a flow rate of 0.8 ml/min; injector and detector temperatures were 270°C.

## GC–MS

Varian 3700 chromatograph with a CP Sil 5CB column, (25 m × 0.25 mm i.d., film thickness 0.39  $\mu$ m) combined with Varian MAT 44S ionization energy 70 eV,

\*Correspondence to: A. Rustaiyan, School of Pharmacy, Shahid Beheshty University of Medical Science, PO Box 14155-6153, Tehran, Iran.

**Table 1.** Composition of the essential oils from *Nepeta ispahamica* Boiss. and *N. binaludensis* Jamzad. (column: 25 m CP Sil 5CB)

Compound	RRI	<i>N. ispahamica</i> %	<i>N. binaludensis</i> %	Identification
$\alpha$ -Thujene	926	–	0.4	GC/MS
$\alpha$ -Pinene	935	3.1	1.1	GC/MS
Sabinene	970	1.9	0.8	GC/MS
$\beta$ -Pinene	976	10.7	3.2	GC/MS, <sup>1</sup> H-NMR
Dehydro-1,8-cineole	986	–	0.5	GC/MS
<i>p</i> -Cymene	1018	–	2.7	GC/MS
1,8-Cineole	1027	65.2	42.3	GC/MS, <sup>1</sup> H-NMR
$\beta$ -Phellandrene	1032	–	0.8	GC, GC/MS
<i>trans</i> -Sabinene hydrate	1059	0.6	0.4	GC, GC/MS
Linalol oxide (furanoid)	1065	–	0.2	GC/MS
<i>cis</i> -Sabinene hydrate	1088	0.4	–	GC, GC/MS
Linalol	1091	–	4.0	GC/MS
<i>trans</i> -Pinocarveol	1129	1.2	–	GC/MS
Verbenol	1134	0.5	–	GC/MS
Pinocarvone	1144	0.8	–	GC/MS
$\delta$ -Terpineol	1153	1.0	3.0	GC/MS
Terpinen-4-ol	1167	1.0	2.8	GC, GC/MS
Myrtenal	1175	1.0	–	GC/MS
$\alpha$ -Terpineol	1177	2.0	4.0	GC, GC/MS
Myrtenol	1184	1.0	–	GC/MS
4 $\alpha$ ,7 $\alpha$ ,7 $\alpha$ -Nepetalactone	1324	–	0.7	GC/MS
4 $\alpha$ ,7 $\alpha$ ,7 $\alpha$ -Nepetalactone	1337	–	25.2	GC/MS, <sup>1</sup> H-NMR
$\beta$ -Caryophyllene	1422	0.2	–	GC/MS
$\beta$ -Caryophyllene oxide	1575	2.1	–	GC/MS

carrier gas He and injector temperature 270°C. Approximately 0.1  $\mu$ l of neat oil was injected under split conditions (100:1) and the oven temperature was held at 60°C for 5 min, programmed at 5°C/min to 220°C and then held at this temperature for 20 min.

## Results and Discussion

The essential oil was obtained from the air-dried parts of *N. ispahamica* in 0.2% yield; that from *N. binaludensis* in 0.8% yield. The identification of the compounds was carried out by comparison of their MS and/or <sup>1</sup>H-NMR spectra (Table 1) with those of authentic samples together with the relative retention indices (RRI). Only the compounds representing at least 0.1% of the mixture are given in the Table in order of their elution on the CP Sil 5CB column. Altogether, 24 constituents, accounting for 92.7% and 91.6% of the oils respectively, were identified. The results shown in Table 1 reveal a clear difference in the chemical composition of the oils studied.

While the oil obtained from *N. binaludensis* contains the monoterpenes usually found in the *Nepeta* oils (1,8-cineole 42.3%, 4 $\alpha$ ,7 $\alpha$ ,7 $\alpha$ -nepetalactone, 25.2%; and 4 $\alpha$ ,7 $\alpha$ ,7 $\alpha$ -nepetalactone, 0.75%) the proportion of monoterpenes in the essential oil of *N. ispahamica* is quite different. In this oil we could not find any trace of nepetalactone or its stereoisomers. On the other hand the amount of 1,8-cineole was higher (65.2%) than that found in the oil of *N. binaludensis*. In the oils isolated

from other *Nepeta* species, the percentage of 1,8-cineole varied from traces to 80%.<sup>4,8–11</sup>

Recently the chemical constituents of the essential oils of *Nepeta italica* L. and *Nepeta sulfuriflora* P. H. Davis from Turkey have been described.<sup>10</sup> Both contain 1,8-cineole (80.8% and 61.5%, respectively). Concerning other significant monoterpenes, both samples are qualitatively similar, although they contain different constituents that characterize the *Nepeta* oils. The lack of nepetalactones is known in the oils of some *Nepeta* species; *N. leucophyl*,<sup>12</sup> *N. discolor*<sup>13</sup> (Himalayan species), *N. italica* from Turkey<sup>10</sup> and *N. cataria* L. cv. *citriodora* growing wild in the Drome region of France.<sup>11</sup> Concerning the sesquiterpenes, the essential oil of *N. ispahamica* contained  $\beta$ -caryophyllene (0.24%) and  $\beta$ -caryophyllene oxide (2.16%) that did not occur in the oil *N. binaludensis*. In the oil of this species we were not able to identify any sesquiterpenes.

The sesquiterpene region (RRI 1375–1980) showed only some trace constituents which could not be identified.

*Acknowledgements* — The authors are grateful to Professor P. Weyerstahl Institute of Organic Chemistry, Technical University of Berlin for GC, GC–MS and <sup>1</sup>H-NMR spectra and Mr V. Mozafarian for helpful assistance in collecting plant specimens and for botanical identification.

## References

1. K. H. Rechinger, *Nepeta*, In *Flora Iranica, Labiatae*, No. 150, eds. K. H. Rechinger and I. C. Hedge, Akademische Druck und Verlagsanstalt, Graz, Austria (1982).

2. A. Zargari, *Medicinal Plants*, 4th edn, Vol. 4. Tehran University Publications, (1990).
3. C. Bourrel, F. Perineau, G. Michel and J. M. Bessiere, *J. Essent. Oil Res.*, **5**, 159 (1993).
4. H. L. De Pooter, B. Nicolai, L. F. De Buyck, P. Goetghebeur and N. M. Schamp, *Phytochemistry*, **26**, 2311 (1987).
5. O. P. Agarwal, D. S. Khanna and R. B. Arora, *Artery*, 487 (1978).
6. A. T. Bottini, V. Dev, D. J. Garfagnoli, H. Lohani, C. S. Mathela and A. K. Pant, *Phytochemistry*, **26**, 1200 (1987).
7. K. Sahurai, K. Jkeda and K. Mori, *Agric. Biol. Chem.*, **52**, 2369 (1988).
8. H. L. De Pooter, B. Nicolai, J. De Laet, L. F. De Buyck, N. M. Schamp and P. Goeghebeur, *Flavour Fragr. J.*, **3**, 155 (1988).
9. S. Pavlovic, A. L. Sevarda, G. A. Kuznetsova, P. Zivanovic, R. Jancic and S. Vujcic, *Arh. Pharm.*, **35**, 227 (1985).
10. G. Kökdil, S. Kurucu and G. Topçu, *Flavour Fragr. J.*, **12**, 33 (1997).
11. J. C. Chalchat and J. Lamy, *J. Essent. Oil Res.*, **9**, 527 (1997).
12. A. T. Bottini, V. Dev, G. C. Shah, C. S. Mathela, A. B. Melkani, A. T. Nerio and N. S. Sturm, *Phytochemistry*, **31**, 1653 (1992).
13. C. S. Mathela, H. Kharkwal and R. Laurent, *J. Essent. Oil Res.*, **6**, 519 (1994).